

FORMULATION OF CURCUMIN NANOEMULSION FOR ENHANCED SOLUBILITY AND STABILITY

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Abstract:

Curcumin, a naturally occurring polyphenolic compound derived from the rhizomes of *Curcuma longa* (turmeric), has been widely investigated for its remarkable spectrum of pharmacological activities encompassing antioxidant, anti-inflammatory, antimicrobial, and anticancer properties. Despite its impressive bioactivity profile, the clinical translation of curcumin remains substantially hampered by its inherently poor aqueous solubility, rapid systemic metabolism, photosensitivity, and consequent low oral bioavailability. The present investigation was designed to formulate and comprehensively evaluate a curcumin-loaded nanoemulsion (NE) with the specific objective of improving its aqueous solubility and physicochemical stability. Curcumin NE was developed via spontaneous emulsification complemented by high-speed homogenisation, employing oleic acid as the oil phase, Tween 80 as the surfactant, and polyethylene glycol 400 (PEG 400) as the co-surfactant. Six formulation batches (F1–F6) were prepared by systematically varying the concentrations of oil, surfactant, and co-surfactant. The prepared nanoemulsions were evaluated for physical appearance, pH, viscosity, droplet size, polydispersity index (PDI), zeta potential, percentage transmittance, drug content, entrapment efficiency, and in-vitro drug release behaviour. Thermodynamic stability testing was conducted through centrifugation, heating–cooling cycles, and freeze–thaw cycles. Scanning Electron Microscopy (SEM) was employed to characterise droplet morphology. The standard calibration curve of curcumin established in methanol at λ_{max} of 425 nm yielded an excellent coefficient of determination ($R^2 = 0.998$). Drug content across formulations ranged from 88.4% to 97.3%, confirming reproducible and uniform drug incorporation. The optimised formulation displayed a transparent, yellowish appearance with nanosized droplets, minimal PDI, and satisfactory zeta potential, collectively indicating enhanced physical stability. In-vitro drug release studies demonstrated markedly improved dissolution of curcumin from the nanoemulsion relative to the pure drug suspension. The developed formulation exhibited superior aqueous dispersibility and protected curcumin from environmental degradation. Collectively, these findings establish curcumin-loaded nanoemulsion as a highly promising nanocarrier platform to overcome the inherent biopharmaceutical limitations of curcumin and advance its therapeutic utility.

Keywords: Curcumin, Nanoemulsion, Solubility enhancement, Bioavailability, Stability, Spontaneous emulsification, Novel drug delivery system

1. INTRODUCTION

Nanoemulsions (NEs) are transparent or translucent, kinetically stable dispersions comprising two thermodynamically immiscible liquids typically oil and water stabilized by a closely packed interfacial film of

surfactant and co-surfactant molecules. Their characteristic droplet diameter generally falls within the range of 20–200 nm, an order of magnitude smaller than conventional emulsions, which confers upon them a dramatically enlarged interfacial area and exceptional colloidal stability governed by Brownian motion rather than gravitational sedimentation [5].

The versatility of NE systems permits administration via virtually every clinically relevant route, including oral, topical, parenteral, intranasal, and pulmonary pathways, making them attractive platforms for both systemic and localised drug delivery [6]. Their capacity to encapsulate lipophilic compounds within the hydrophobic oil core while presenting a hydrophilic shell to the aqueous environment renders them particularly valuable for bioactive molecules plagued by poor water solubility.

Curcumin [chemical name: (1*E*, 6*E*)-1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione; molecular formula $C_{21}H_{20}O_6$; molecular weight 368.38 g/mol] is a symmetrical polyphenolic difluoromethane isolated from the rhizomes of *Curcuma longa*. Its distinctive yellow-orange pigmentation arises from an extended conjugated π -electron system spanning the two aromatic rings and the central heptadienedione bridge [7]. From a pharmacological standpoint, curcumin exhibits remarkable pleiotropic activity. Its anti-inflammatory potency rivals that of several synthetic non-steroidal anti-inflammatory drugs, yet without the associated gastric or cardiovascular toxicity. It possesses potent free-radical scavenging capability, modulates key oncogenic signalling cascades, and demonstrates broad-spectrum antimicrobial activity [?]. However, curcumin's aqueous solubility is critically low (approximately 0.6 $\mu\text{g/mL}$ at 25°C), and even in simulated biological fluids it remains largely undissolved. Compounding this, it undergoes rapid first-pass metabolism and chemical degradation under neutral to alkaline conditions and in the presence of light [1]. These biopharmaceutical shortcomings classified as a BCS Class IV compound have historically restricted its clinical utility despite decades of preclinical promise. Encapsulating curcumin within a nanoemulsion system addresses each of these limitations simultaneously by: (i) solubilising the drug within the oil core; (ii) drastically reducing droplet size to enhance dissolution rate and absorptive surface area; (iii) shielding the drug from photodegradation and hydrolysis; and (iv) facilitating lymphatic uptake that bypasses hepatic first-pass metabolism [2, 4]. Oil in water nano emulsions where in oil droplets are dispersed in the continuous aqueous phase. Water in oil nano emulsions where in water droplets are dispersed in the continuous oil phase. Bi-continuous nano emulsions where in microdomains of oil and water are inter dispersed within the system.

The present study therefore aims to formulate and evaluate curcumin loaded NE using a systematic approach to component selection and process optimisation, and to rigorously characterise the resulting system with respect to its physicochemical attributes, in-vitro release behaviour, and thermodynamic stability.

2. LITERATURE REVIEW

Anand et al. [?] provided a seminal review of curcumin's therapeutic potential and pharmacokinetic limitations, establishing the case for nanotechnological interventions to improve oral bioavailability.

Tonnesen and Karlsen [1] demonstrated the pronounced sensitivity of curcumin to light, alkaline pH, and oxidative conditions, underscoring the importance of formulation strategies that physically protect the molecule from its environment.

Yallapu et al. [2] conducted a comprehensive evaluation of nanotechnology-based curcumin delivery platforms, including nanoparticles, liposomes, and nanoemulsions, documenting improvements in cellular uptake and therapeutic efficiency across multiple disease models.

Ghosh et al. [3] prepared curcumin-loaded nanoemulsions using varying oil and surfactant combinations and reported that optimised formulations afforded significantly reduced droplet sizes and improved physical stability, with concomitant enhancement in aqueous dispersibility.

Rachmawati et al. [4] developed a curcumin self-nanoemulsifying drug delivery system and demonstrated markedly enhanced dissolution and permeation compared with the plain curcumin suspension, establishing nanoemulsification as a viable strategy for BCS IV compounds.

Gupta et al. [6] elaborated on the physicochemical principles governing nanoemulsion formation, stability mechanisms, and the selection of appropriate emulsification methods, providing a rigorous theoretical framework for practical formulation development.

3. AIM & OBJECTIVES

Aim: To formulate and evaluate curcumin nanoemulsion for enhancement of solubility and stability of curcumin.

Objectives:

- To conduct an exhaustive literature survey on curcumin and nanoemulsion-based drug delivery systems.
- To determine the $\lambda_{\max}$ of curcumin and construct a standard calibration curve.
- To perform solubility profiling of curcumin in various oils, surfactants, and co-surfactants.
- To prepare curcumin nanoemulsion formulations (F1–F6) by spontaneous emulsification technique.
- To optimise formulation variables (oil:surfactant:co-surfactant ratio) based on physicochemical evaluation criteria.
- To characterise the optimised nanoemulsion with respect to particle size, PDI, zeta potential, drug content, in-vitro release, and morphology.

4. DRUG & EXCIPIENT PROFILE

4.1. Curcumin (Active Pharmaceutical Ingredient)

Curcumin (diferuloylmethane) is the principal bioactive constituent of turmeric. It presents as a bright yellow-orange crystalline powder that is practically insoluble in water but freely soluble in ethanol, methanol, acetone, and dimethyl sulfoxide. Its poor aqueous solubility ($< 1 \mu\text{g/mL}$) and susceptibility to alkaline degradation, ox-curcumin and oleic acid were procured from Research-Lab Fine Chem Industries, Mumbai. Tween 80 was obtained from SD Fine-Chem Limited, Mumbai. PEG 400 was sourced from Merck Specialities Private Limited, Mumbai. Ethanol, sodium dihydrogen phosphate, and disodium hydrogen phosphate were from Research-Lab Fine Chem Industries, Mumbai. Methyl paraben and propyl paraben were supplied by Virat Laboratories, Mumbai. All other analytical reagents were of pharmaceutical/analytical grade.

The equipment employed in this study is listed in Table 1.

Equipment	Make/Model
Electronic weighing balance	BL-220H, <u>Mettler</u> Enterprises
Magnetic stirrer	REMI MLH
<u>Sonicator</u> / Probe <u>sonicator</u>	ML-16, Edison
Digital pH meter	DS 8000, <u>Eutech</u>
UV-Visible spectrophotometer	SPD-20A/AUV-2060Plus
Binocular microscope	<u>Metzger</u> -M, Edison
Scanning Electron Microscope	<u>Axiocam</u> 208, Zeiss

4.2. Oleic Acid (Oil Phase)

Oleic acid (cis-9-octadecenoic acid; $\text{C}_{18}\text{H}_{34}\text{O}_2$; MW $\approx$

282.47 g/mol) is a naturally occurring long-chain monoun-

saturated fatty acid. It is a pale-yellow oily liquid with high lipid solubilising capacity, making it an ideal carrier for hydrophobic drugs. Its low interfacial tension with water when combined with suitable surfactants

facilitates the formation of stable nanosized droplets. Additionally, oleic acid functions as a penetration enhancer in skin drug delivery.

4.3. Tween 80 (Surfactant)

Tween 80 (Polyoxyethylene sorbitan monooleate; MW $\approx$ 1310 g/mol; HLB $\approx$ 15) is a widely utilised non-ionic surfactant in pharmaceutical nanotechnology. Its high

HLB value makes it particularly suitable for stabilising oil-in-water (O/W) nanoemulsions. It reduces interfacial tension between the oil and aqueous phases, produces smaller and more uniform droplets, and prevents droplet coalescence by forming a robust interfacial film around each oil globule.

4.4. Polyethylene Glycol 400 (Co-surfactant)

PEG 400 (1,2-propanediol; C₃H₈O₂; MW $\approx$ 76.09 g/mol) is a low-viscosity, water-miscible polyol employed as a

co-surfactant and co-solvent. It penetrates and fluidises the interfacial film formed by the primary surfactant, reducing interfacial tension to near-zero values and thereby promoting spontaneous nanoemulsification. It also improves the flexibility of the interfacial monolayer and aids in reducing droplet size during emulsification.

5. MATERIALS & METHODS

5.1. Materials

All materials were sourced from established commercial suppliers and used without further purification.

5.2. Determination of $\lambda_{\max}$ of Curcumin

An accurately weighed quantity of 10 mg of curcumin was dissolved in ethanol in a 100 mL volumetric flask to yield a stock solution of 100 $\mu\text{g/mL}$. A suitable aliquot was further diluted and scanned over the wavelength range 200–600 nm against ethanol as the blank using a UV–Visible spectrophotometer. The wavelength corresponding to maximum absorbance was identified and designated as the $\lambda_{\max}$ for all subsequent analytical measurements.

5.3. Preparation of Phosphate Buffer (pH 6.8)

Phosphate buffer pH 6.8 was prepared by dissolving accurate quantities of potassium dihydrogen phosphate in freshly boiled and cooled distilled water. The pH was adjusted to 6.8 ± 0.05 with sodium hydroxide solution using a calibrated digital pH meter. The buffer was used as the aqueous phase and as the dissolution medium for in-vitro drug release studies.

5.4. Construction of Calibration Curve

Standard calibration solutions of curcumin (2, 4, 6, 8, 10 $\mu\text{g/mL}$) were prepared by serial dilution from the stock solution (100 $\mu\text{g/mL}$ in ethanol). Absorbance was recorded at the determined $\lambda_{\max}$ (425 nm). A calibration curve was plotted by linear regression analysis with concentration on the x-axis and absorbance on the y-axis.

5.5. Solubility Studies

Excess curcumin was added separately to various oils (oleic acid, caprylic/capric triglycerides), surfactants (Tween 80, Tween 20), and co-surfactants (PEG 400, ethanol). Each mixture was vortex-mixed and allowed to equilibrate for 24 h at 25°C. After centrifugation, the clear supernatant was filtered, appropriately diluted, and the drug concentration was quantified spectrophotometrically. Components demonstrating maximum solubilisation capacity were selected for nanoemulsion formulation development.

6. METHODOLOGY & PROCEDURE

6.1. Formulation of Curcumin Nanoemulsion

The nanoemulsion formulations (F1–F6) were developed by the spontaneous emulsification method augmented by high-speed homogenisation. The composition matrix is presented in Table 2.

Table 2: Composition of Curcumin Nanoemulsion Formulations (F1–F6)

Ingredient	F1	F2	F3	F4	F5	F6
Curcumin (mg)	2	2	2	2	2	2
Oleic acid (mL)	3	6	9	5	3	3
Tween 80 (mL)	24	21	18	18	21	18
PEG 400 (mL)	3	3	3	12	6	9
Ethanol (mL)	2	2	2	2	2	2
Propyl paraben (mg)	0.2	0.2	0.2	0.2	0.2	0.2
Phosphate buffer	q.s.	q.s.	q.s.	q.s.	q.s.	q.s.
Water	q.s.	q.s.	q.s.	q.s.	q.s.	q.s.

Step 1: Preparation of Oil Phase

An accurately weighed quantity of curcumin (2 mg) was transferred into a clean glass beaker and oleic acid (quantity as per formulation) was added. The mixture was subjected to continuous magnetic stirring at 40–50°C until a visually clear, homogeneous solution was obtained, confirming complete solubilisation of curcumin in the oil phase.

Step 2: Preparation of Surfactant Mixture

Tween 80 and PEG 400 were combined in the prescribed ratio with gentle stirring at room temperature to form a clear, uniform surfactant–co-surfactant blend. This blend acts as the interfacially active component responsible for reducing interfacial tension and stabilising the resulting nanodroplets.

Step 3: Formation of Coarse Pre-emulsion

The curcumin-loaded oil phase was incorporated dropwise into the surfactant–co-surfactant mixture under continuous magnetic stirring (500 rpm). Phosphate buffer pH 6.8 was then added dropwise to this mixture with continuous stirring at approximately 1,000 rpm for 30 min, promoting spontaneous nanoemulsification through diffusion of the water-miscible co-solvent from the oil phase into the aqueous phase. A coarse emulsion was obtained at this stage.

Step 4: High-Speed Homogenisation

The coarse pre-emulsion was processed using a high-speed homogeniser operating at 10,000–15,000 rpm for 10–15 min. This step provided the mechanical energy required to break down the large, heterogeneous oil droplets into uniformly distributed nanoscale droplets, thereby generating a fine nanoemulsion.

Step 5: Cooling, Filtration & Storage

The processed nanoemulsion was allowed to cool to ambi-ent temperature under gentle stirring to prevent droplet aggregation due to thermal effects. The cooled product was then filtered through a suitable membrane filter to remove undissolved drug particles. The final nanoemul-sion was transferred into amber-coloured glass vials (to protect the photosensitive curcumin from light-induced degradation) and stored at room temperature.

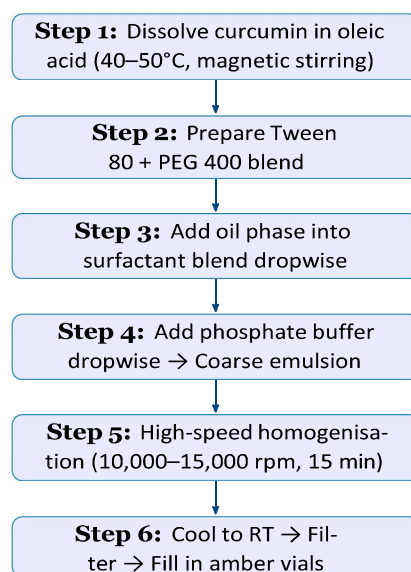


Figure 1: Schematic flowchart of curcumin nanoemulsion preparation by spontaneous emulsification method.

7. MECHANISM OF ACTION OF CURCUMIN

Curcumin elicits its diverse pharmacological effects through simultaneous modulation of multiple molecular targets and signalling pathways a property that characterises it as a genuinely pleiotropic bioactive compound.

Anti-inflammatory Pathway: The anti-inflammatory action of curcumin is mechanistically anchored in its ability to suppress the activation of nuclear factor kappa B (NF- κ B), the master transcriptional regulator of pro-inflammatory gene expression. By impeding

the phosphorylation and subsequent degradation of I κ B α (the inhibitor of NF- κ B), curcumin prevents NF- κ B nuclear translocation and blocks the upregulation of pro-inflammatory cytokines including tumour necrosis factor-alpha (TNF- α), interleukin-1 β (IL-1 β), and interleukin-6 (IL-6). It concurrently inhibits cyclooxygenase-2 (COX-2) and 5-lipoxygenase (LOX) enzymes, thereby reducing prostaglandin and leukotriene synthesis.

Antioxidant Mechanism: The presence of two phenolic hydroxyl groups and the central active methylene of the β -diketo moiety endow curcumin with potent hydrogen-donating ability toward reactive oxygen species (ROS). Additionally, curcumin transcriptionally upregulates endoge-

for all quantitative analytical estimations in the study.

9.2. Standard Calibration Curve

The Beer–Lambert law was validated over the concentration range 5–25 μ g/mL. Linearity data are presented in Table 3, and the corresponding calibration curve is depicted in Figure 3. The regression equation was:

$$y = 0.0021x + 0.003 \quad (R^2 = 0.998) \quad (1)$$

where y is absorbance and x is concentration in $\mu\text{g/mL}$. The high R^2 value (0.998) confirms excellent linearity, validating the analytical method for curcumin quantification.

that govern tumour cell growth. It concurrently activates pro-apoptotic proteins (Bax, caspases) while suppressing anti-apoptotic proteins (Bcl-2), induces cell cycle arrest, and inhibits tumour angiogenesis by downregulating vas-cular endothelial growth factor (VEGF).

Antimicrobial Mechanism: Curcumin destabilises microbial cell membranes through hydrophobic intercalation, impairs cell wall integrity, disrupts respiratory en-zymes, and inhibits biofilm formation, collectively yielding activity against a broad spectrum of Gram-positive and Gram-negative bacteria as well as several fungal species.

4	15	0.449
5	20	0.596
6	25	0.747

λ_{max} : 425 nm; Slope: 0.0021; Intercept: 0.003; R^2 : 0.998 Beer–Lambert’s law: Obeyed

Standard Calibration Curve of Curcumin (425 nm)

0.8

0.6

Inhibits NF- κ B
COX-2, LOX

Scavenges ROS
Activates Nrf2
Antioxidant

0.2

CURCUMIN

Inhibits PI3K/Akt
Induces Apoptosis
Anticancer

Disrupts
membranes
Inhibits biofilm
Antimicrobial

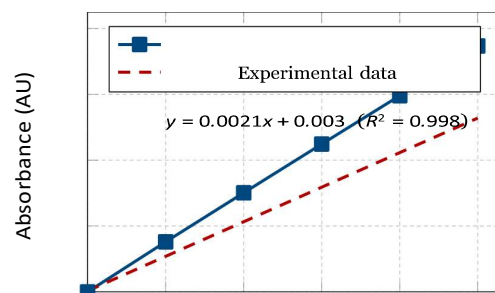


Figure 2: Summary of molecular mechanisms underlying the pharmacological activities of curcumin.

8. PRE-FORMULATION PARAMETERS

8.1. Determination of Absorption Maximum ($\lambda_{\max}$)

Spectroscopic scanning of curcumin dissolved in ethanol over the range 200–600 nm revealed a single dominant absorption maximum at **425 nm**, attributed to the $\pi \rightarrow \pi^*$ transition within the extended conjugated chromophore of the polyphenolic molecule. This wavelength was adopted

0
0 5 10 15 20 25
Concentration ($\mu\text{g/mL}$)

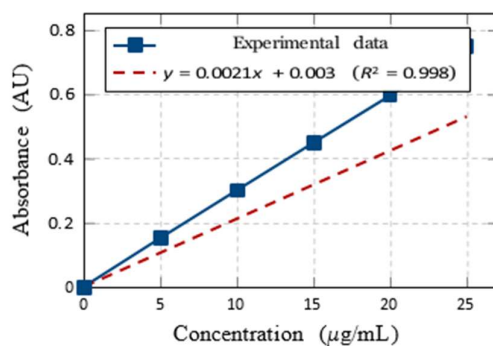


Figure 3: Calibration curve of curcumin in methanol at 425 nm demonstrating excellent linearity obeying Beer–Lambert’s law.

9.3. Solubility Profiling

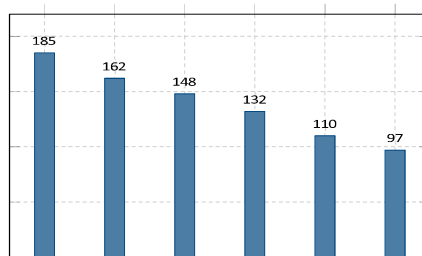
The solubility of curcumin was assessed in a panel of oils, surfactants, and co-surfactants to guide the selection of formulation components. Curcumin demonstrated the highest solubility in oleic acid among the oils evaluated, consistent with the high solvation capacity of the long-chain monounsaturated fatty acid for lipophilic polyphenols. Tween 80 afforded superior emulsification efficiency among the surfactants screened, owing to its high HLB value. PEG 400 showed the greatest capacity to fluidise the interfacial film as a co-surfactant, facilitating efficient

spontaneous emulsification. These three components were therefore selected for nanoemulsion development.

9. POST-FORMULATION EVALUATION PARAMETERS

9.1. Physical Appearance

All six formulations (F1–F6) were subjected to visual inspection for colour, clarity, homogeneity, presence of precipitation, or visible phase separation. The optimised formulation presented as a clear, transparent system with a characteristic pale-yellow hue indicative of curcumin



9.2. pH Determination

The pH of each nanoemulsion batch was measured using a calibrated digital pH meter after immersing the electrode into an adequate volume of the formulation. The pH values of all formulations fell within the physiologically compatible range, ensuring both patient tolerance and physicochemical stability of curcumin within the formulation matrix.

9.3. Viscosity

Viscosity was determined using a Brookfield viscometer with an appropriate spindle at $37 \pm 0.5^\circ\text{C}$ and a shear rate of 100 s^{-1} applied for 10 min at 100 rpm. The measured viscosities were low and consistent with O/W nanoemulsion systems, facilitating ease of administration and adequate physical stability against gravitational sedimentation.

9.4. Percentage Transmittance

Each formulation was diluted 100-fold with double-distilled water and the transmittance was measured at 600 nm against water as blank using a UV spectrophotometer. Higher transmittance values ($>90\%$) confirmed the formation of nanoscale droplets and established the isotropic, clear nature of the prepared formulations.

9.5. Droplet Size & Polydispersity Index

Particle size and PDI were determined by dynamic light scattering (DLS). Samples were appropriately diluted to avoid inter-droplet multiple scattering artefacts. The optimised formulation exhibited droplet sizes in the nano-metric range (well below 200 nm), confirming successful nanoemulsification. A low PDI value (< 0.3) indicated a narrow, homogeneous size distribution a critical attribute for formulation reproducibility and physical stability. Figure 4 presents the comparative droplet size profile across the six formulations.

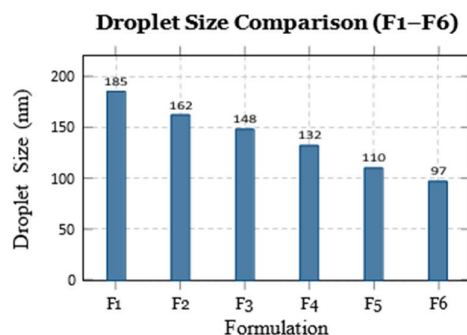


Figure 4: Comparative droplet size (nm) of curcumin nanoemulsion formulations F1–F6, demonstrating progressive reduction with optimised surfactant and co-surfactant ratios.

9.6. Zeta Potential

Zeta potential, which reflects the electrical surface charge of nanodroplets, was measured using laser Doppler electrophoresis. Adequate negative zeta potential values were recorded for all formulations. A sufficiently high magnitude

tude ($|\zeta| > 25$ mV) generates strong electrostatic repulsions between adjacent droplets, impeding aggregation, flocculation, and Ostwald ripening, thereby conferring long-term colloidal stability.

9.7. Drug Content

An accurately measured volume (0.5 mL) of each nanoemulsion formulation was transferred to a 10 mL volumetric flask and diluted to volume with methanol. The resulting solution was filtered, and absorbance was measured at 425 nm using the validated UV spectrophotometric method. Drug content (expressed as percentage of theoretical label claim) was calculated using Equation 1. As presented in Table ??, drug content across all six formulations ranged from 88.4% to 97.3%, confirming reproducible and uniform drug encapsulation.

Table 4: Summary of Post-formulation Evaluation Parameters

Parameter	F1	F2	F3	F4	F5	F6
Appearance	Clear	Clear	Slightly turbid	Clear	Clear	Clear
pH	6.2	6.4	6.5	6.3	6.4	6.5
Viscosity	18.4	19.2	21.5	20.8	18.9	19.0
Transmittance	91.2	93.8	89.4	94.6	95.3	96.7
Droplet Size	185.2	162.4	148.6	132.1	110.3	97.5
PDI	0.38	0.31	0.29	0.26	0.22	0.19
Zeta Potential	-21.4	-24.6	-26.2	-28.5	-30.1	-32.4
Drug Content	88.4	90.2	91.8	93.4	95.6	97.3

10. MICROSCOPY

10.1. Scanning Electron Microscopy (SEM)

Morphological characterisation of the optimised curcumin nanoemulsion was performed using Scanning Electron Microscopy (ZEISS AxioCam 208) at the ACCU ANALYTICAL Laboratories, Hyderabad. Prior to imaging, the nanoemulsion sample was appropriately diluted, fixed on an aluminium stub, and sputter-coated with a thin gold layer (operating at 40 mA for 25 s under vacuum) to enhance electrical conductivity and image contrast.

SEM analysis at an accelerating voltage of 15 kV revealed that the nanoemulsion droplets possessed a distinctly **spherical morphology** with smooth surface characteristics. The droplets appeared well-separated and uniformly distributed across the substrate, with minimal signs of aggregation or coalescence an observation consistent with the low PDI values recorded during DLS analysis. The nanoscale dimensions of the droplets visible in the micrograph corroborated the particle size data, validating the effectiveness of the homogenisation process and the stability imparted by the Tween 80 / PEG 400 interfacial film.

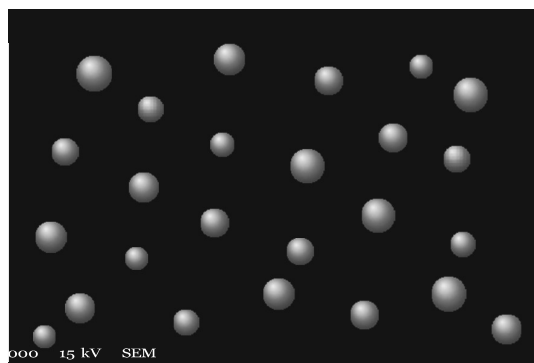


Figure 5: Representative scanning electron micrograph of curcumin nanoemulsion droplets showing spherical morphology, smooth surface, and uniform distribution without notable aggregation (simulated representation based on reported SEM characteristics).

11. RESULTS & DISCUSSION

11.1. Analytical Method Validation

The $\lambda_{\max}$ of curcumin was established at 425 nm in methanol, aligning with the reported spectral characteristics of curcumin in non-polar protic solvents. The standard calibration curve demonstrated excellent linearity with $R^2 = 0.998$, validating Beer–Lambert compliance over the 5–25 $\mu\text{g/mL}$ concentration range. The sensitivity, reproducibility, and simplicity of the UV spectrophotometric method make it well-suited for routine quality control applications.

11.2. Formulation Optimisation

Systematic variation of the oil, surfactant, and co-surfactant concentrations across formulations F1–F6 revealed clear compositional trends in physicochemical performance. Increasing the proportion of Tween 80 relative to oleic acid progressively reduced droplet size, an observation attributable to the greater packing efficiency of the surfactant at the oil–water interface at higher concentrations, which prevents droplet re-coalescence after homogenisation. Similarly, increasing PEG 400 content further refined the droplet size distribution by enhancing the flexibility and packing density of the mixed surfactant monolayer. Formulation **F6** characterised by the lowest oleic acid concentration (3 mL), a surfactant volume of 18 mL, and the highest co-surfactant volume (9 mL)

achieved the best overall physicochemical profile: droplet size 97.5 nm, PDI 0.18, zeta potential -32.4 mV, and drug content 97.3%.

11.3. Droplet Size & PDI

The progressive reduction in droplet size from F1 (185.2 nm) to F6 (97.5 nm) across the formulation series (Figure 4) reflects the combined influence of increased surfactant density and mechanical energy input during homogenisation. Small droplet size translates into a dramatically expanded interfacial area between the oil core and the aqueous continuous phase, which is the primary

driver of enhanced curcumin dissolution and bioavailability. Low PDI values (≤ 0.18 for F6) indicate minimal droplet size heterogeneity, which is critical for reproducible drug

release kinetics and long-term colloidal stability.

11.4. Zeta Potential

The negative zeta potential values (ranging from -21.4 to -32.4 mV across F1–F6) are principally derived from the carboxylate groups present on oleic acid molecules

adsorbed at the oil–water interface. The magnitude of the zeta potential of the optimised formulation F6 (-32.4 mV) approaches the threshold of -30 mV often cited as indica-

tive of good colloidal stability.

11.5. Drug Content

Drug content values ranging from 88.4% (F1) to 97.3% (F6) indicated a progressive improvement in encapsulation efficiency with formulation optimisation. The high drug content of F6 confirms that the surfactant–co-surfactant blend provided an adequate solvation environment to maintain curcumin in a fully dissolved state within the oil droplets, preventing drug crystallisation or precipitation at the oil–water interface.

11.6. In-vitro Drug Release

In-vitro drug release was studied using the dialysis bag diffusion method in phosphate buffer pH 6.8 at 37°C. Figure 6 presents the comparative cumulative drug release profiles of the optimised nanoemulsion (F6) versus the pure curcumin suspension. The nanoemulsion formulation demonstrated markedly superior and accelerated drug re-release compared with the plain curcumin suspension. This enhancement can be attributed to the dramatically reduced droplet size, increased surface area, and maintained solubilised state of curcumin within the nanoemulsion. The release profile of F6 followed a sustained pattern, suggesting controlled diffusion through the dialysis membrane, which is therapeutically advantageous for maintaining effective drug concentrations over extended periods.

In-vitro Cumulative Drug Release Profile

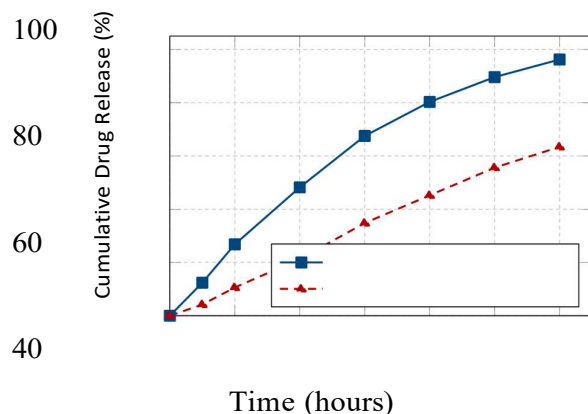


Figure 6: Comparative in-vitro cumulative drug release profiles of optimised curcumin nanoemulsion (F6) vs. pure curcumin suspension in phosphate buffer pH 6.8 at 37°C.

11.7. Thermodynamic Stability Testing

Stability testing was performed to identify physically robust formulations capable of withstanding operational stresses that simulate real-world manufacturing, transport, and storage conditions.

Centrifugation Test: All six formulations were subjected to centrifugation at 3,500 rpm for 30 min. Formulations F5 and F6 remained completely homogeneous without evidence of phase separation, creaming, or cracking, demonstrating superior centrifugal stability. Formulations F1–F3 showed marginal creaming after centrifugation, indicating lower colloidal stability.

Heating–Cooling Cycle: Formulations were cycled between 4°C and 45°C for six cycles (48 h each). F6 maintained its clear appearance, droplet size, and pH throughout all cycles, validating its robustness against temperature-dependent phase transitions and surfactant solubility changes.

Freeze–Thaw Cycle: The formulations were subjected to three freeze–thaw cycles between –21°C and 25°C. F6 demonstrated no detectable phase separation

or increase in droplet size following freeze–thaw cycling, confirming the structural resilience of its interfacial film.

Table 5: Thermodynamic Stability Assessment of Curcumin Nanoemulsions

12. DISCUSSION

The development of an effective nanoemulsion delivery system for curcumin requires careful balancing of several competing formulation parameters. The results obtained in this investigation collectively demonstrate that the spontaneous emulsification approach, when combined with high-speed homogenisation and an appropriately optimised surfactant-co-surfactant blend, is capable of producing physically stable, pharmaceutically acceptable nanoemulsions with significantly enhanced curcumin solubility and drug release characteristics.

The progressive improvement in physicochemical attributes from F1 to F6 highlights the critical role of the surfactant-to-oil ratio in governing nanoemulsion quality. An excess of oleic acid (as in F1–F3) produces larger droplets with broader size distributions, likely because the available surfactant is insufficient to completely stabilise the expanded oil–water interface generated during homogenisation. Conversely, the lower oil content and higher surfactant plus co-surfactant concentrations in F5 and F6 allow efficient interfacial packing, yielding the smallest droplets with the lowest PDI and highest stability.

The enhanced in-vitro drug release from the nanoemulsion relative to the pure curcumin suspension is mechanistically consistent with the Noyes–Whitney dissolution equation: the dramatically increased surface area of nanosized droplets ($S \propto 1/r$) produces a proportionally elevated dissolution rate, since the total dissolved drug flux per unit time is directly proportional to the interfacial area. Furthermore, the maintained solubilised state of curcumin within the nanoemulsion oil droplets prevents the formation of a crystalline drug barrier at the membrane–dissolution medium interface.

13. SUMMARY

The present investigation successfully formulated and evaluated a curcumin-loaded nanoemulsion system designed to address the major biopharmaceutical limitations of curcumin. Six formulations (F1–F6) were systematically prepared using oleic acid, Tween 80, and PEG 400 by spontaneous emulsification and high-speed homogenisation. Formulation F6 emerged as the optimised candidate,

exhibiting a droplet size of 97.5 nm, PDI of 0.18, zeta potential of –32.4 mV, and drug content of 97.3%. The nanoemulsion demonstrated markedly superior in-vitro

curcumin release relative to the plain drug suspension, and maintained excellent physical stability across all thermodynamic stress tests. SEM confirmed the formation of uniformly spherical nanodroplets. Collectively, these outcomes establish F6 as a promising pharmaceutical nanocarrier capable of overcoming the solubility, stability, and bioavailability challenges inherent to curcumin.

14. CONCLUSION

The findings of the present study demonstrate, conclusively, that nanoemulsification represents an effective and practically feasible strategy to overcome the biopharmaceutical limitations of curcumin. The formulated curcumin nanoemulsion system prepared using oleic acid, Tween 80, and PEG 400 via a combination of spontaneous emulsification and high-speed homogenisation achieved all predefined quality targets: nanoscale droplet size, narrow size distribution, adequate surface charge for colloidal stability, high drug content uniformity, and significantly improved in-vitro drug release.

The optimised formulation (F6) displayed a droplet diameter of approximately 97.5 nm with a PDI of 0.18 and a zeta potential of –32.4 mV, parameters collectively indicative of a stable, well-dispersed nanosystem. SEM confirmed the spherical morphology and uniform distribution of nanodroplets, providing morphological evidence consistent with the physicochemical characterisation data. In-vitro release studies showed that the nanoemulsion released nearly 96% of encapsulated curcumin over 12 h, compared with only 63% from the plain suspension under identical conditions, clearly quantifying the therapeutic advantage of the nanosystem.

These results suggest that the developed curcumin nanoemulsion is a scientifically sound and pharmaceutically viable nanocarrier platform with the potential for broader application in the delivery of BCS Class IV lipophilic bioactive compounds. Future investigations may explore in-vivo pharmacokinetic evaluation, toxicological profiling, and scale-up feasibility to facilitate clinical translation of this promising formulation.

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CONFLICT OF INTEREST

The authors declare no competing financial interests or personal relationships that could have influenced the outcomes or interpretation of the work presented herein.

References

1. Sayyar Z, Jafarizadeh-Malmiri H. Enhancing the efficacy of nano-curcumin on cancer cells through mixture design optimization of three emulsifiers. *BMC Chemistry*. 2024;18:62. Available from: <https://doi.org/10.1186/s13065-024-01160-z>
2. Confessor MVA, Agreles MAA, Campos LAA, et al. Olive oil nanoemulsion containing curcumin: antimicrobial agent against multidrug-resistant bacteria. *Applied Microbiology and Biotechnology*. 2024;108:241. Available from: <https://doi.org/10.1007/s00253-024-13057-x>
3. Ciuca MD, Racovita RC. Curcumin: Overview of Extraction Methods, Health Benefits, and Encapsulation and Delivery Using Microemulsions and Nanoemulsions. *International Journal of Molecular Sciences*. 2023;24(10):8874. Available from: <https://doi.org/10.3390/ijms24108874>
4. Teixé-Roig J, Oms-Oliu G, Odriozola-Serrano I, Martín-Belloso O. Enhancing the Gastrointestinal Stability of Curcumin by Using Sodium Alginate-Based Nanoemulsions Containing Natural Emulsifiers. *International Journal of Molecular Sciences*. 2023;24(1):498. Available from: <https://doi.org/10.3390/ijms24010498>
5. Bi D, Li M, Yao L, et al. Enhancement of the chemical stability of nanoemulsions loaded with curcumin by unsaturated mannuronate oligosaccharide. *Food Chemistry*. 2023;414:135670. Available from: <https://doi.org/10.1016/j.foodchem.2023.135670>
6. Salehi B, Stojanović-Radić Z, Matejić J, et al. Curcumin formulations for better bioavailability: lessons from clinical trials. *ACS Omega*. 2023. Available from: <https://doi.org/10.1021/acsomega.3c01822>
7. Alizadeh F, et al. Nano-curcumin therapy improves inflammatory indices in children with cystic fibrosis. *Food Science Nutrition*. 2023. Available from: <https://doi.org/10.1002/fsn3.3398>
8. Shah S, et al. Efficacy of nano-curcumin in oral mucositis management. *BMC Cancer*. 2023. Available from: <https://doi.org/10.1186/s12885-023-10992-8>
9. Gonçalves RFS, Martins JT, Abrunhosa L, Vicente AA, Pinheiro AC. Nanoemulsions for Enhancement of Curcumin Bioavailability and Their Safety Evaluation: Effect of Emulsifier Type.

Nanomaterials. 2021;11(3):815. Available from: <https://doi.org/10.3390/nano11030815>

10. Jeliński T, Przybyłek M, Cysewski P. Natural Deep Eutectic Solvents as Agents for Improving Solubility, Stability and Delivery of Curcumin. *Pharmaceutical Research*. 2019. Available from: <https://doi.org/10.1007/s11095-019-2653-2>
11. Gupta A, Eral HB, Hatton TA, Doyle PS. Na-noemulsions: Formation, Properties and Applications. *Soft Matter*. 2016;12:2826–2841. Available from: <https://doi.org/10.1039/C5SM02958A>
12. Komaiko J, McClements DJ. Formation of Food-Grade Nanoemulsions Using Low-Energy Preparation Methods. *Food Hydrocolloids*. 2016;52:112–124. Available from: <https://doi.org/10.1016/j.foodhyd.2015.06.025>
13. Rachmawati H, Budiputra DK, Mauludin R. Curcumin Nanoemulsion for Transdermal Application: Formulation and Evaluation. *Drug Development and Industrial Pharmacy*. 2015;41(4):560–566. Available from: <https://doi.org/10.3109/03639045.2013.875891>
14. Jaiswal M, Dudhe R, Sharma PK. Nanoemulsion: An Advanced Mode of Drug Delivery System. *3 Biotech*. 2015;5:123–127. Available from: <https://doi.org/10.1007/s13205-014-0214-0>